

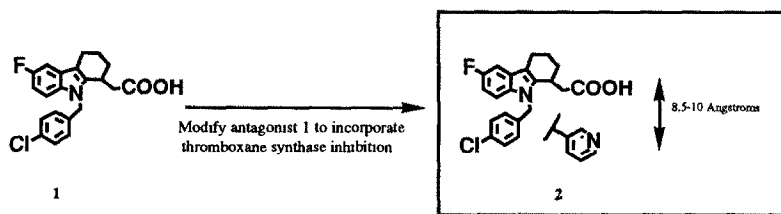
K. Russell<sup>§</sup>, H. Gaskin, R. Jessup  
Research Department, ICI Pharmaceuticals, Alderley Park, Macclesfield, Cheshire, SK10 4TG, England  
<sup>§</sup>Current address, Medicinal Chemistry Department, ICI Pharmaceuticals Group, ICI Americas Inc.,  
Wilmington, DE 19897

**Abstract:** A modification of a known thromboxane antagonist by the incorporation of a tethered pyridine moiety which gave a series of dual acting thromboxane antagonist/thromboxane synthase inhibitors is described. Compound 9 inhibits human thromboxane synthase with an  $IC_{50}$  of 50nM and is an antagonist of human platelet aggregation with a  $pA_2$  of 6.0. This compound also demonstrates *ex vivo* activity when dosed orally to rats at 10mg/kg. p.o.

A strategy which combines the potential benefits from both of the above uses a dual acting thromboxane antagonist/thromboxane synthase inhibitor.<sup>5</sup> Such a compound would be expected to increase the levels of  $\text{PGI}_2$ , as well as other beneficial prostaglandins such as  $\text{PGD}_2$  and  $\text{PGE}_2$ , by virtue of redirection of the biosynthetic route (Scheme 1) and decrease the levels of  $\text{TXA}_2$ . Elevated levels of  $\text{PGH}_2$ , which is itself a thromboxane agonist, are blocked by the antagonist activity of the compound. Studies in animals and in normal human volunteers with mixtures of  $\text{TXA}_2$  antagonists and  $\text{TXA}_2$  synthase inhibitors have demonstrated that the two agents have greater therapeutic benefits when given in combination than when given individually.<sup>6</sup>

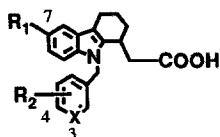


We describe herein our search for such a dual acting agent by the structure modification of a known TXA<sub>2</sub> antagonist (1).<sup>7</sup>



Our approach was based on the idea that it might be possible to find a way of modifying 1 which would incorporate thromboxane synthase inhibition without losing thromboxane antagonist activity. Heme-binding heterocycles such as pyridine or imidazole are reported to confer good inhibitory activity for thromboxane synthase when combined with a carboxylic acid using an appropriate linker.<sup>8</sup> It has also been shown previously that the heterocyclic nitrogen to carboxylate distance was in the range of 8.5 - 10 angstroms in the most active thromboxane synthase inhibitor.<sup>9</sup> It was suggested that the 8.5 - 10 angstrom distance represented the Fe-Heme to carboxylate binding site separation in the enzyme. Recognizing that 1 contains a carboxylic acid we decided to explore the incorporation of a pyridine unit into 1 in the hope that it might be possible to pick up thromboxane synthase inhibitory properties and not lose thromboxane antagonist activity. We used the geometric information cited above to direct our initial efforts to find a suitable position for incorporation of the pyridyl substituent in the tetrahydrocarbazole nucleus. Initial attempts to replace the benzylic group in 1 with a 3-pyridylmethyl moiety led to compound 3 (Table 1) which was only weakly active as a thromboxane antagonist, and was inactive opposite thromboxane synthase. Since a 8.5 - 10 angstrom distance between the carboxylate and the pyridine nitrogen could not be attained in 3, we then tried extension of the carboxylate chain to attain this separation between the functional groups. This gave a compound, which was found to be devoid of either thromboxane antagonist or thromboxane synthase inhibition.

We next turned to replacement of the 7-fluoro substituent with pyridine containing side chains. Although substitution at this position gave compounds which showed weak thromboxane synthase inhibition we were discouraged by the loss of thromboxane antagonist activity. In fact simple replacement of the 7-fluoro substituent in 1 with methoxy led to an order of magnitude decrease in the thromboxane antagonist activity of this molecule. Thromboxane antagonist activity is clearly sensitive to changes at this site.



Compound	R <sub>1</sub> <sup>*</sup>	R <sub>2</sub> <sup>*</sup>	X	Thromboxane antagonist (pA <sub>2</sub> ) <sup>#</sup>	Thromboxane synthase inhibition (IC <sub>50</sub> , μM) <sup>*</sup>
1	F	Cl	H	7.1	NA <sup>-</sup>
3	F	H	N	5.5	NA
4	F	3-[-CH <sub>2</sub> CH <sub>2</sub> (CH <sub>2</sub> ) <sub>3</sub> Py]	C-H	5.4	0.5
5	F	3-[-OCH <sub>2</sub> (CH <sub>2</sub> ) <sub>3</sub> Py]	C-H	6.3	0.1
6	F	3-[-O(CH <sub>2</sub> ) <sub>5</sub> -Py]	C-H	5.5	0.3
7	F	4-[-O(CH <sub>2</sub> ) <sub>3</sub> -≡-Py]	C-H	6.8	0.6
8	F	4-[-O(CH <sub>2</sub> ) <sub>4</sub> -≡-Py]	C-H	6.7	0.4
9	F	3-[-O(CH <sub>2</sub> ) <sub>3</sub> -≡-Py]	C-H	6.0	0.05
10	F	3-[-O(CH <sub>2</sub> ) <sub>4</sub> -≡-Py]	C-H	5.6	0.04
11	F	3-[-O(CH <sub>2</sub> ) <sub>3</sub> -≡-Py]	C-H	6.2	0.2

<sup>\*</sup> Py = 3-pyridyl. <sup>-</sup> NA = not active. <sup>#</sup> Thromboxane antagonist potency against U 44619 induced platelet aggregation using human platelet rich plasma. <sup>\*</sup> Thromboxane synthase inhibition using human microsomal TXA<sub>2</sub> synthase; results are expressed as IC<sub>50</sub>s for the inhibition of TxB<sub>2</sub> formation from <sup>14</sup>C labelled arachidonic acid.<sup>13</sup>

Table 1

Surprisingly substitution on the chlorobenzyl moiety proved to be more profitable. The activity of a series of compounds substituted in the 3- or 4- position of the phenyl group are shown in Fig. 1 and Table 1. It can be seen that in general thromboxane antagonist potency is retained with both substitution patterns although the most potent compounds are found with the pyridine - containing substituent in the 4- position. Inhibiting thromboxane synthase, however, appears to be more effective in compounds with the pyridine - containing substituent in the 3- position, with chain lengths of 4 to 5 (Fig 1).

Of course these compounds are all very flexible and they must pay an entropic penalty for this when they bind to the enzyme or receptor in terms of the loss of internal conformational freedom.<sup>10</sup> However, for each methylene unit in the sidechain we gain an increment of binding energy by virtue of an increased hydrophobic effect.<sup>11</sup> These two binding energy components may largely cancel out. The advantage of having such a flexible linking unit is that the terminal pyridine unit can more easily adopt the correct three dimensional position relative to the rest of the molecule. Therefore, we viewed the linker groups as not contributing to the binding energy in a major way but merely serving as a means of holding the pyridine moiety so that it could bind to the heme iron center. Thus these initial studies gave us important information relating to the feasibility of achieving good TXA<sub>2</sub> synthase inhibition while maintaining TXA<sub>2</sub> antagonism. Our strategy from this point was to try to reduce the conformational flexibility of the side chains by the incorporation of geometrically rigid elements in the hope that greater potency would result. We were aware,

however, that the conformation adopted by the compound when it binds to the enzyme may be different from that which binds to the receptor.

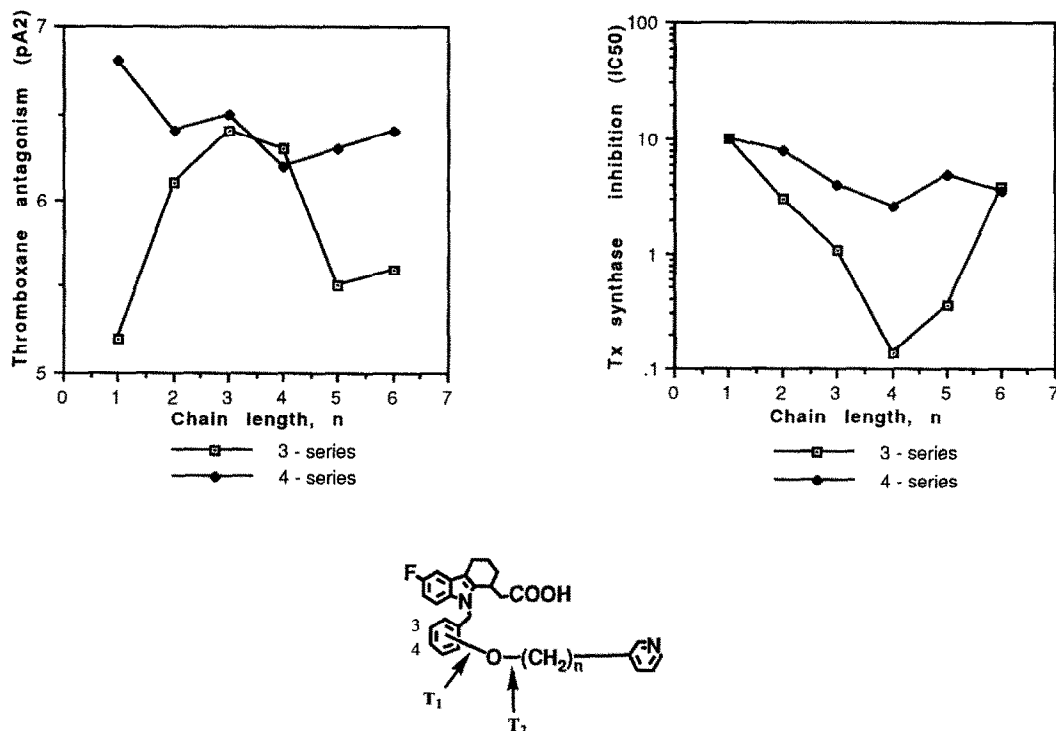
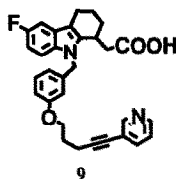


Fig 1

The phenolic link in the side chain is thought to introduce one element of rigidity with T<sub>1</sub> and T<sub>2</sub> preferring torsions of 0 degrees and 180 degrees respectively.<sup>12</sup> Replacement of the CH<sub>2</sub>-CH<sub>2</sub> unit with the less conformationally flexible O-CH<sub>2</sub> leads to an increase in both activities (see 4 and 5). Incorporation of an acetylene unit in place of a CH<sub>2</sub>-CH<sub>2</sub> in 6 provided another increase in potency to give 9. In the 4-substituted series compounds 7 and 8 showed good antagonist activity and a reasonable level of thromboxane synthase inhibition. Potent thromboxane synthase inhibition is seen in the 3-series with 9 and 10 with the former being a reasonably good antagonist. Reduction of the acetylene yielded the *cis*-olefin 11 which although slightly more potent than 9 was a poorer enzyme inhibitor.



Compound **9** was found to exhibit the dual properties of thromboxane synthase inhibition and receptor antagonism. When dosed orally (10mg/kg) to conscious rats compound **9** caused inhibition (86%, 77%, and 71%) of platelet TXA<sub>2</sub> receptors at 1, 3 and 5 hours after dosing, respectively.<sup>13</sup>

In conclusion we have demonstrated the successful modification of a known TXA<sub>2</sub> antagonist structure to obtain a dual action compound which involved appending a heme-binding pyridyl moiety. The synthesis and testing of a series of flexibly linked pyridines allowed us to evaluate the feasibility of the general approach. Increased potency was obtained by incorporation of an acetylene unit which restricted the conformational flexibility of the linking chain. The SAR to date has demonstrated that it is difficult to match high antagonist potency with potent TXA<sub>2</sub> inhibition in this system and may be suggestive of different conformational requirements for the TXA<sub>2</sub> receptor and TXA<sub>2</sub> synthase. Oral activity was demonstrated by **9** in an *ex vivo* rat model.<sup>13</sup>

#### References and Notes

1. a) Svensson, J., Hamberg, M., Samuelsson, B. *Proc. Natl. Acad. Sci. U.S.A.* **98**, 285-294 (1976).  
 b) Hamberg, M., Hedqvist, P., Strandberg, K., Svensson, J., Samuelsson, B. *Life Sci.* **16**, 451-461 (1975).  
 c) Samuelsson, B., Folco, G., Grantstrom, E., Kindahl, H., Malmsten, C., *Advances in Prostaglandin and Thromboxane Research*, Cocceani, F., Olley, P. M., Eds., Raven. New York: 1978, Vol. 4, pp 1-26.
2. a) Lefer, A. M., *Drug News and Perspectives*, **2**, 265 (1989).  
 b) Ogeltree, M. L. *Fed. Proc., Fed., Am. Soc. Exp. Biol.*, **46**, 133 (1987).  
 c) Lefer, A. M., Darius, H., *Ibid.*, **46**, 144 (1987).  
 d) Halushka, P., Mais, D. E., Saussy, D. L., Jr., *Ibid.*, **46**, 149 (1987).
3. a) Collington, E. W., Finch, H., *Annu. Rep. Med. Chem.*, **25**, 99-108 (1989).  
 b) Wolanin, D. J., Campbell, J. B., *Annu. Rep. Med. Chem.*, **26**, 113-122 (1991).
4. Witt, W., Muller, B., *Advances in Prostaglandin and Thromboxane Research*, Samuelsson, B., Paoletti, R., Ramwell, P. W., Eds., Raven. New York: 1987, pp 279-284.
5. a) Gresele, P., Deckmyn, H., Nenci, G. G., Vermeylen, J., *Trends Pharmacol. Sci.*, **12**, 158-163 (1991).  
 b) Bhagwat, S. S., Gude, C., Cohen, D. S., Lee, W., Furness, P., Clarke, F. H., *J. Med. Chem.* **34**, 1790-1797 (1991).
6. a) Fitzgerald, D. J., Fragetta, J., Fitzgerald, G. A., *J. Clin. Invest.* **82**, 1708 (1988).  
 b) Shebuski, R. J., *Circulation*, **76**, IV-101 (1987).  
 c) Gresele, P., Arnout, J., Deckmyn, H., Huybrechts, E., Pieters, G., Vermeylen, J., *J. Clin. Invest.*, **80**, 1435 (1987).
7. a) Girard, Y., Yoakim-Rancourt, P., Hamel, P., Gillard, Guindon, Y., J. W., Letts, G., Evans, J., Leveille, Ethier, D., Lord, A., Jones, T. Masson, P., Ford-Hutchinson, A. W., Rokash, J., *Prog. Clin. Biol. Res.*, **301**, 585-589 (1989).  
 b) See accompanying paper in this journal for a description of the synthetic approaches used to access compounds described here.
8. a) Tanouchi, T., Kawamura, M., Ohyama, I., Kajiwara, I., Iguchi, Y., Okada, T., Taniguchi, K., Miyamoto, T., Hayashi, M., *J. Med. Chem.*, **24**, 1149-1155 (1981).  
 b) Cross, P. E., Dickinson, R. P., Parry, M. J., Randall, M. J., *J. Med. Chem.*, **28**, 1427 (1985).  
 c) Cross, P. E., Dickinson, R. P., Parry, M. J., Randall, M. J., *J. Med. Chem.*, **29**, 342-346 (1986).

- d) Ford, N. F., Browne, L. J., Cambell, T., Gemenden, C., Goldstein, R., Gude, C., Wasley, W. F., *J. Med. Chem.*, **28**, 164 (1985).
- e) Kato, K., Ohkawa, S., Terao, S., Terashita, Z., Nishizaki, K., *J. Med. Chem.*, **28**, 287 (1985).
9. Iizuka, K., Akahane, K., Momose, D., Nakazawa, M., Tanouchi, T., Kawamura, M., Ohyama, I., Kajiwara, I., Iguchi, Y., Okada, T., Taniguchi, K., Miyamoto, T., Hayashi, M., *J. Med. Chem.*, **24**, 1139 (1981).
10. Estimated to be ca. 0.9 kcal/mol per  $sp^3$ - $sp^3$  rotation. Page, M. I., Jencks, W. P., *Proc. Natl. Acad. Sci. U.S.A.*, **68**, 1678-1683 (1971).
11. The incremental Gibbs energy of transfer per methylene unit from octanol to water is ca. 0.7 kcal/mol. See Hansch, C. Coats, E., *J. Pharm. Sci.*, **59**, 731 (1970). From protein folding studies the free energy of transfer of a methylene from water to the hydrophobic core of the protein was found to be favourable by 1.0 - 1.6 kcal/mol. The discrepancy between these two numbers arises because two surfaces are buried when a protein folds - both the methylene group and the portions of the protein to which it packs. See Kellis J. T., Nyberg, K., Fersht, A. R., *Biochemistry*, **28**, 4914-4922 (1989).
12. Spellmeyer, D. C., Grootenhuis, P. D. J., Miller, M. D., Kuyper, L.F., Kollman, P. A., *J. Phys. Chem.*, **94**, 4483-4491 (1990).
13. Thromboxane antagonism was assessed using a blood platelet aggregation test based on that described by Born [*Nature*, **194**, 927-929 (1962)]. Human citrated, platelet-rich plasma was aggregated by the addition of the  $TXA_2$  mimetic agent U46619 so that a dose- response curve was generated. This was repeated in the presence of increasing amounts of test compound (generally in the range  $10^{-5}M$  to  $10^{-10}M$ ) and a  $K_b$  was calculated. An apparent  $pA_2$ , indicating antagonist potency, was thus generated (where  $pA_2 = -\log K_b$ ). In *ex vivo* rat experiments antagonism was measured as a dose ratio (that is the ratio of the concentration of U46619 required to cause a 50% of platelet aggregation in the presence and absence of the test compound)
- Thromboxane synthase inhibition was demonstrated using the standard *in vitro* test procedure used by Howarth [*Biochem. Soc. Transactions*, **10**, 239-240 (1982)] using a human platelet microsomal  $TXA_2$  synthase preparation and using a quantitative thin layer radiochromatographic method to assess the conversion of [ $1-^{14}C$ ] arachidonic acid to the  $TXA_2$  metabolite thromboxane  $B_2$ .